

Fla⁻ H₁^{non b} segregants.

1256

DATE: APR 28 1955

REF:

	1	2	3	4	5	6	7	8	9	10
1	1252A2	{	a-x b							
2	"									
3	1252B2		tails? in b serum.							
Test by two regimes. Isolate Fla ⁻ H ₁ ^x										
B ₂)	x	FA10 (H ₁ ^b)	+ b serum	→	?	Fla ⁺ H ₁ ^b				
A ₂)	x	FA93 H ₁ ^a	+ a serum.	If x = a, no swarms. If x = b, H ₁ ^x = b.						
Results:										
1	-	Not pure motile	Give to DCC							
2	-	Pure motile	. 3 { B no swarms							
3	-	" "	. 3 { A swarms →							

DATE: MAY 3 1955

REF:

A. Staining in situ W1177.

a. in mixture c: 0.005% T₂ under oil - No
stg. overnight Some inhibition?b. In methanol 4000 20% (vs Penassay) + c: .10% T₂,
isolated colonies only (resistant?) - sterilized in center.? are these conditions too aerobic? Otherwise polynitrate toxicity
of T₂.Should re-isolate colonies; compare growth c, s T₂ under oil.

B. Chemis from isolated cells. (see c. 4-6 May then ref.)

1. In scattered region many chemis showed intestinal lethals.
(effect of cold?)

2. Few & granules now seen.

3. Isolates immediately spread growth under them (film of
mortality from dego 1) A-B-C

D 2 is tangle

D 4 c 20 cells, 7 term.

5 4 g.

6 4 g.

7 4 g.

10

X

E 1. tangle, 2 term.

2-5 n.s.

6 no 2

7 no 2

8 no 2

9 4 g.

40

50

Z chains. etc.
Motility in methanol solns.

1208

DATE: MAY 4 1955

REF:

- A. attempts at leaving cells in situ, according in relation to 28u HMM beads, in Methanol 4000cps 2% in benzene. General conclusions as stated attached. see 1208 p. 100.
- 10 ^{Some} ~~ethanol~~ ^{noted} ~~probably~~ ^{more frequent} ~~after refrigeration~~ ^{but this} ~~is not settled by direct observation~~
- B. 5/6/55. Set up to repeat 1254. Began OK but slow to divide at RT (though warm) & later lost c.g.
- C. " Methanol 400 cps seems to slow up motile cells (129A1+). Put. trials as selector for E cells. But must. used 4000 in trials.
- D. 30 See 5/8/55. E - a slump: did not completely inhibit initial nucle. up-take.
- 40 1) Possible use of Scotchbrite HMM beads as reform markers - there is a slow drift; May be better to use 3% Methanol 4000 rather than 2%.
- 2) Wasted exp. in counting E, Z chains
- 3) Pulvin exp. on screening E, S cells by viscous medium extremely successful. See ff.
- 50 4) But most time last week or 10 days was wasted exp. to improve general impression & technique.

5/6/00 1258

Lab plans: what to do? Things are a mess.

1. Currently enmeshed in the fate of Z granules. Can these really give any important information? By following a granule during the growth of a single cell, one might get a clue as to whether growth is interstitial or bipolar (in a few cases). To distinguish, one might have to show increasing separation between two granules, before fission in a single cell and this may be difficult.

It is already clear that 1) terminal granules usually remain terminal, and that this is the most common type, already suggesting a polarity in the cell. Occasionally, bi-antipolar cells are seen (more commonly than bi-synpolar), suggesting that the two poles share something distinct from the fissile center. However, the basic interest in the Z granule for the current problem is the possible correlation with E, and this, if anything is what should be pursued for now. Later it may be convenient to try to repeat experiments with a polar-flagellated organism.

Another sideline is to use the chains in stiff medium to study other problems, chiefly lethality both spontaneous and UV. Also look for data on growth of branched cells. (Twort)

2. More pertinent: 1) look for divided E further. 2) diagnose E, S cells by viscous media. 3) transfer intermediate chains for electron microscopy 4) clean up serotypes of co-segregants-- collect more? 5) For 4 and others need to complete review of data and write up.

3. TODAY: Clean up what is accumulated to look at and photograph.

Start new preps. of 93--x w/wo TZ. Use for divided clones and for Z correlation.

(Sat 5/7/55- Sun 5/8/55—)

Use T2 stained prepn. 5/6. 12n7 Checked first with 1237A1+ for swarm motility. In this series, used 2% methocel 400, diluted c. 1/10 with penassay.

a) use methocel for trap; b) isolate initials in broth trap, then TRANSFER to mcl.

The latter was found ineffective (probably still too stiff); By 4 PM, had isolated 13 cells still sluggishly motile in mcl trap, and 7 addl. which were at a distance from reservoir but not now motile. swarm cells were sluggishly motile in this methocel conc., about 50-70% were directly inhibited. This oln. probably wets glass more effectively, at any rate it tends to spread, and a few of the motiles below may be contaminants from 1237A1+.

The motile residuals above were ^{planted} in individual drops of broth for class. as Sw. or E cells.

NR found, in first group: 6 swarms, 3 E, 2 ng, 2 E.
second 4 E 1 ng 2 E

Total 6 S 7E 3ng 4 E

which demonstrates strong selection against E cells

Detailed counts:

	growth	motiles
1.	4+	9
2	3 = sw	swarm, 50%?
3	4+	2
4	ng	
5	4+	12 (from Z cell, but Z nf)
6	map 4+	0,1
7	sn, 1 mot cell	
8	20 sw	16, sev. shakes, prob. sw
9	sw sw	
10	sw sw	
11	like 8 sw	
12	500	12
13	200 sw sw	
21	4+	4
22	ng	
23	4+	18
24	4+	24
25	4+	2
26	4+	15
27	4+	16

(104)

The occasion was also used to plant about 25 single motiles (removed before test below—perhaps should have been left in it) for ~~opn~~ ^{opn} ~~co~~ ^{co} ~~st~~ st ~~ance~~ ^{ance} on immediate and later motility of dividing chain cell. About 12 usable cases no discrepancies, some to one or two later divisions. As none gave two motiles, pres, none of these were E. Of remainder, most gave two app. nm at this division—it may be possible to reexamine these drops tomorrow. What is significance of this crisis in termination? Is is growth in fresh medium? (May still need a good exhausted medium to keep cell size small.)

P8 These were then used in tests for residual motility in mcl. Unf., 1,5 were wasted in 5% mcl 15 (calc. visc 200) which proved also to inh. swarms. Further tests were then made with mcl 400, 1.8% and 1% (1:1 penassay), the latter being adopted as it permits almost full motility of motile swarms (from above). (This may be too fluid for accurate discrimination against E, as will be seen). From E: 12, 23, 24, 26, 27, ~~altogether~~ cells reisolated which remained motile were planted for further classification → none proved definite E cells. See further below.

Until this is worked out against in further pedigree!

-x SW 666 initials in methocel
and Z chains
b a serum.

1259

DATE: May 9, 1955

REF:

1	2	3	4	5	6	7	8	9	10
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Prestained prepn. used. (st Z 9:45-12N; phage to 1:20, centr. and refr. 1:40)
(also another preph. unstained - A.

Note immoderate spreading out of methocel droplets. isolate initials in 1% methocel 400
(1:1 2% penassay).

A. isol. from unstained, plant out in droplets individually.

B. isol. Z-stained initials. to c. 3PM, some fresh isol. C to 4:30
me t in single drops on initial cg. first, transfer latter as families to
isolation cg. Ditto for A-- plant out descendants.

D. 5PM B above, in broth traps: pick c. 4000 initials (somewhat late now for tests)
in serums.

(Klein visited 5/10-11.)

D: Almost all initials are inhibited in a or b serum, though cells may continue to spin
for a few minutes. 7 cells did persist in b, planted out. 3 proved viable swarms.
isolate as 1259 D1-3. See DCG for results of platings(after picking to broth)in
MGA. 2 persists in a, but neither viable.

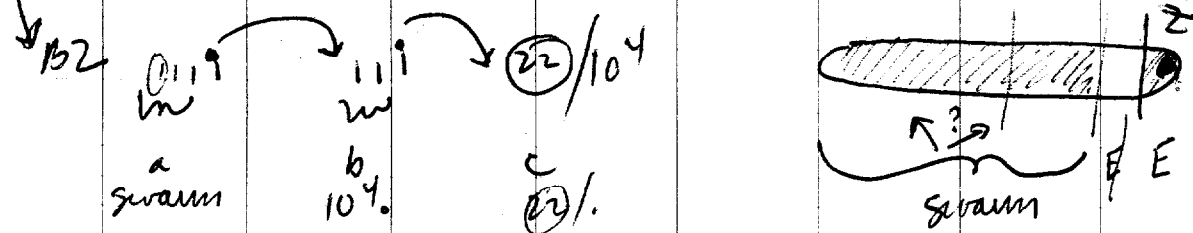
E. same as B-D but not sublined

A: Held to 5/11 for examin, and may have partly diminished therefore.

13 clones 2? E clones. 1% Methocel probably too thin..

B: Most isolates grew out; had been separated once or twice at n_2-n_4 . However, of
38 isolations, 3 ng; 5 swarms; only 4E, none interesting except:
E-clones were reexamined for content. In B2, sib to swarms had 22 motiles,
transferred to 238/ E2,3; 4; 5(sw). The motiles in E2 tested, all gave rise to
inviable or E clones & therefore certainly not sw. cells/

6% originally looked as if only c. $100+ / 10^4$ but these later proved to be swarms.
The clone was not recovered (owing to drying out) to verify original low assay.



Otherwise, detailed numbers of intermediates were not recorded.
F+ 5/10 interm motiles tested in a serum: at least (28) from 5 clones were immo.
but 2/4 from B15 were not. However, two tested swarms were inhibited; specificity
of serum should be rechecked.

Also saved 1259B1 (= b8). Swarm- test purity by plating
B2a, B (= c5 z cell removed at n_5 = nonmot, b) (not not certain record)

DCG found D1-3 all motile but with confusing clusters. B1: no definite swarms B2b
"all clusters"; a pure non-motile. Will have to be rechecked on return

E: 34 isolates planted w/o lineage afterward.

~~AE~~ (9,11,15,16)
~~15E~~ (1, 4,3,1,1,6,7,3,3,5,5,2,1,3,1,4,....)
~~4~~ Sw
5 ng.

Only conclusion: medium not adequately selective. Try 1½% methocel 400
(v.i.: 1260)

1259 summary to 5/16

5/12 Plated in MGA

5/13 Picked possible singles. Plates were incubated too short a time at 37° , D1 & D2 had singles, swarms, & clusters; D3, B1, & B2 & singles & clusters only. Counts:

	<u>Clusters & swarms</u>	<u>Singles</u>	<u>Singles picked</u>	
D1	51	3	2	} these spotted on MGA
D2	91	1	1	
D3	59	5	4	
B1	32	21	8	
B2b	90	17	8	



5/14 All "singles" picked 5/13 & spotted on MGA were motile (Spots had appearance of "clusters" rather than swarms).

Plated again: All original broths, + ^{some of} singles picked 5/13 (D1, 2; D2, 1; D3, 2; B1, 2; B2b, 2.)

Incubated 3 hrs at 37° , overnight at 22° , then refrigerated until examined 5/16.

5/16 Results of 5/14 platings:

Original broths:

- D1 Swarms, centered swarms, & col. c "satellites"  
- D2 ~ D1, higher proportion of swarms.
- D3 ~ D1.
- B1 trails, clusters, apparent singles; no swarms
- B2a pure non-motile
- B2b All clusters

Presumed Fla - :

- D1(1) all clusters
- D1(2) Clusters, swarms } no singles
- D2 Clusters, swarms, no singles
- D3(1) Clusters, swarms } no singles
- D3(2) " " }
- B1(1) } Clusters, trails or satellites; possibly some singles;
- B1(2) } no swarms.

MAY 10 1955

Best resume page

In abc 3 swarms, 5E / 34 isolates

disappointing. No E concl. z. But write out detail any how

B1
 B2

37+ → 15/104
 5/104
 laty 22/104

B10 11 → but only 1 leg seen.

B15 Rec. confused.

c1 11 p →
e2 111 p →
e4 19 →

why 3 drops? (prob Eng.);
i/q →

But only 1 drop, \$100 / 10^4!

~~1000~~ p letters

c5 1 inf \rightarrow /22 \rightarrow where 3 drops? (Ed has 1) prob-
~~12~~
 505

28 11 1111

1500

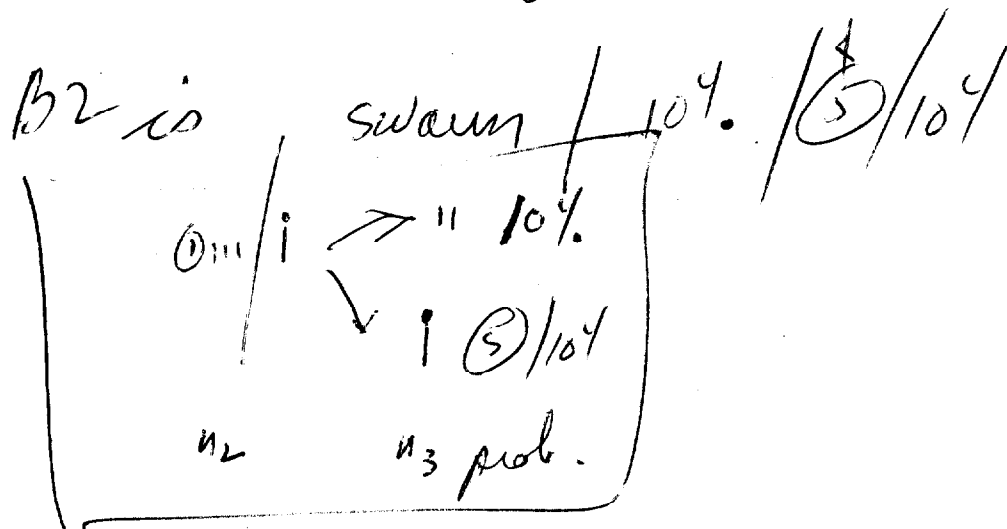
cy / (+) many.
to [238] H2 } Held for full
Residue to F2, F3 } analyses if
needed.

ConcB were evidently confused yesterday!!

MAY 1 0 1955

Do not save these swarms any to possible confusion.

But study closely B2 and C4.



C4 is $0 \rightarrow \frac{57100}{10^4}$.

238 kind before picking

Partitions

May 13. New prepn., unstained. (probably usual, about 90-120mins.)

Fuse drops 2:30 Collect to 3:30. Cf 1259D motile.

No initial was nearly as active as 59D. Pick those that have moved the furthest, not necessarily v. active now. Estimated yield, 10% of broth yield.

Note : to compensate for spreading of methocel solution, use cg. that has been greased (human), then flamed. This works ~~very~~ well, especially with larger drops, but smaller drops are too convex for best visualization. Intention was partly to look for early chains (E) in the methocel, but time did not allow and most isolates were made to broth directly/(A, B resp.) Lineages were separated at n_2 -3.

A: 1,2,3,6 ok. Partitions at n_1 :

14+ :1 6:5 ng snakes. Later transferred entire clones to get fullest estimate of motiles.

A1 came out +(14):6 Sepn at n_1 =

B1-14,21-36. 4 ng. Mostly non E. Records show at first scanning:

2:4;1 14+ 2:1 3 7:20 0:4;1 1:3;d 3 24+ 2:1 1:0 5

sw;sw;sw;sw (1260B33 later DCG verified purity of each). 6:5 7:+

Underscores were rechecked (on ungreaed slide!) and following definite values for splits on these:

1:20 8:20 2:2 4:12 3:2 7: 26 Therefore no equal splits.

General totals:

E 5
ng 4
sw 1
E

33

Little if any selection for E in 1½% methocel.400. Need

2% which probably totally stops many motile cells.

No new experiments after 5/4/
Trip to NY 5/18-5/24/ Reserve lab with
1/3'

→ 82266

1261

Method

JUN 1 1955
MAY 31 1955

1:1
Pupae 93 x 82266, 10⁵ - 11⁴⁵ (12³⁰)
c 430-545 isol. residual motiles. ^{SIC} ^{in antipyrin} Af. c. 4 PM.
Est discrimination factor

A) Note: to prevent spread of methanol, ~~plates~~ ^{c.g.} are lightly greased with fungus (moss side); flamed; oil added. ^{ca 1%} However, motile selection seemed most effective when there was appreciable wetting and spreading of the drops on the coverglass.

Notes transferred to fresh petri dishes c 6 PM, Dec 30^o
Counts of (+) 10² - 10⁴. : 4, 25, 20, 53, 46, 4, 6, 2, 50 ; 3, 20, 18, 3, 20, 7, 11,
13, 10, swarms.

JUN 2 1955

Σ : (6E : 3F : 1 swarm. & 2 Inusable)

\therefore with 2% methanol 400 there is effective discrimination. at this case, Fla⁺ (123701⁺) was greatly slowed down (10x ?) but most cells did continue to move.

[Note - to this point considerable interruption in continuity of work was occasioned by ① trip to NY for ascites meeting ② breakdown of manipulator - valve in diaphragm, temporarily repaired I.

\therefore continue pedigree studies on prescheduled initials.

swarms: manual plating of clone, in 1 ml, .01 ml gave 44 swarms
again note low ratio. Perhaps emulsion! 265 singles

(see photo - plate had been held at RT overnight, inc 2 1/2 hours, then RT 4 hours.

Mcl 400 2% solutions.

1262

Pedigree.

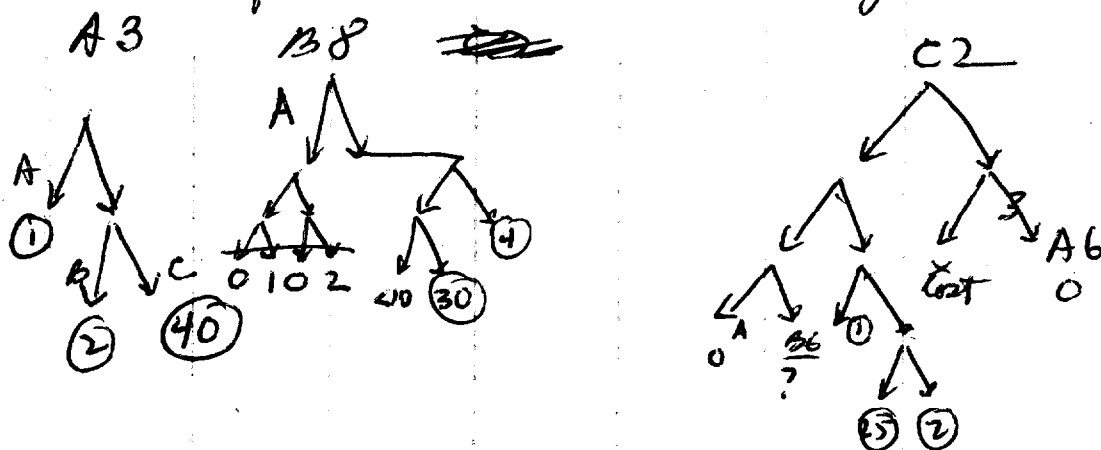
JUN 2 1955

A) grow in penicillin B. grow in Mcl.
Plant to pure drops c. 12¹⁰ PM.

1261 pups.

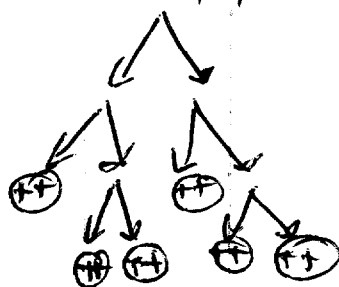
No cells continuously motile like Fla⁺ clone were seen. Pick the most active. If continued, probably used to regulate the degree of wetting. Did ~~pedigree~~ to 43-45 on 34 mutants transferred to broth.

P3, scan for E, ϕ , swarm. Found only 3 E clones.



∴ splits are 1:42 3:34 and 28?:0 (known 1:27)
2:25

One swarm clone C4, already pure. DCC checked purity of each clone by plating.



same! as 1262 C1-5
for H₁ check.

swim totals were only 3E, 1S : 22 ϕ and 10 ϕ (= lethal)
[11-0; 5-1; 2-2, 3, 4, 6, 7],

43, 37, 28

the experiment was quite unsuccessful.

Again review salmonella data
to get paper out of the way.

July 13
1955

A) → Should first get general picture of experiments + what they were!

White out 1138 B4

? → X SW666 $4p^+$

Note diminished motility of large cells. Oct. early isol 1. → $1/10^3$...
(remarks on growth cycle) e.g. 1141 A4

1141 A4 v.p.

A5

B1

B3

B4 vp

A1, A3, B5, C1 n.g. (stayed motile)

C2

Σ	a	b	c	d
3	1	1	5	-
	2	6	6	11
1	0	1	1	1
	0	1	1	1

2 1 2 2 av3.

Σ				
A ₂	1	2	2	19*
B ₂	D _{1/5} 18	6	16	16
	G₂ 15	20	27	27-29
	n.g.			40 45
C ₂	1	2	2	2
B	D _{2/5} 11	9	15	15
D ₂	1	2	2	7
				20
		16	23	33
			23	24-26
				34
				47.

$\Sigma=33$

A4 (v.p.)

* ① first surname!

note partition: 19:11
or 1:(19:11):1:1

But cannot use as G2 may be listed as from B4!

B4 If the latter, then

This datum is unreliable.

subscript = point of this branch in the pedigree.

7/13/55

Σ

1142. (9)

C3

750
5 tested

6 10 31 36

D1

7100
10 tested

7 24 41 45

C4

10
2, 1, 3, 1, 0, 0

3 10 — — *no word*

1143

E3

3

2 2 12 14
3 12 — —

E2

11

s_1	2	2	2	7
s_1	2	2	3	3d
s_2	3	10	16	23
	3	10	42	48
s_7	5	11	27	31
	5	11	31	38
	5	11	14	—
	5	11	—	15

1144

Leifson cultures.

1272
SEP 9 1955

all 6 cultures grow as well or better at 30° as at 37 except 205.

For preliminary comparisons, re-inoculate H1, H302, H32, H37

1:5 in broth + re-inoculate 9AM -

SEP 8 1955

Leifson's slides Helicobacter

figure pairs primarily

H1 A.

1700x apoch.

P. acuminose type

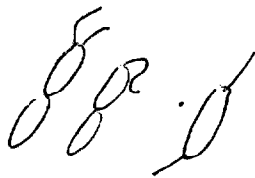


no clear
antipolar
pairs
speaking?



polar mutant
unip.

H 300



ditto
cells larger. than above.

H 32

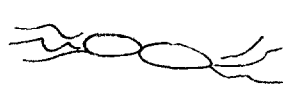


united, antipolar
v. hio?

"alcaligenis"
(*Lyphobacter*)



H 37



large cells.

H 205.



more usual

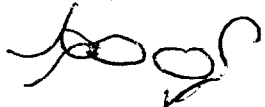


rare

H 430



usually unipolar



prob. intermediate

polar mutants
same bipolar!

(m)

